

Poultry Science Journal

ISSN: 2345-6604 (Print), 2345-6566 (Online) http://psj.gau.ac.ir DOI: 10.22069/psj.2022.20355.1832



Digestible Arginine Requirements in Hy-Line W-36 Laying Hens: Effects on Performance, Egg Characteristics, and Plasma Parameters During 40 to 46 Weeks of Age

Ahmad Azizi¹, Aliasghar Saki¹, Hossein Jahanian Najafabadi¹, Pouya Zamani¹ & Zeinab Bardel¹

Department of Animal Science, Faculty of Agriculture, Bu-Ali Sina University, Hamedan, Iran

Abstract

Poultry Science Journal 2023, 11(1): 103-113

Keywords

Arginine Blood parameters Egg quality Laying hen IGF-1

Corresponding author Aliasghar Saki alisaki34@yahoo.com / asaki@basu.ac.ir

Article history Received: January 06, 2022 Revised: September 21, 2022 Accepted: December 06, 2022 This study was conducted to estimate digestible arginine (Arg) requirements for performance and egg quality parameters in Hy-Line W-36 laying hens from 40 to 46 weeks of age. A total of 150 laying hens were arranged in a completely randomized design with 5 treatments, 5 replicates and 6 hens in each. These treatments included 0.81, 0.86, 0.91, 0.96, and 1.01 % digestible Arg. At the end of experiment, data were obtained in performance, egg quality, and quantity parameters as well as plasma levels of cholesterol, uric acid, globulin, and insulin-like growth factor (IGF-1). Results have shown that feed conversion ratio, egg production, and egg mass improved by supplementing 0.86 digestible Arg in the diet compared to other treatments (P < 0.05). Moreover, these items significantly affected by the interaction of treatments and weeks. In the last three weeks of this experiment, the use of 0.86, 0.91, 0.96, and 1.01 % digestible Arg significantly increased egg mass and egg roduction. Plasma concentrations of globulin, cholesterol, and uric acid were not affected by dietary Arg levels. However, a significant increase in plasma IGF-1 was shown by supplementation of % 0.91 Arg (P < 0.05). Based on quadratic equations, the optimum levels of digestible Arg for egg production, egg mass, feed conversion ratio, and IGF-1 were 0.917, 0.917, 0.908, and 0.970%, respectively.

Introduction

The arginine amino acid (Arg) is an important substrate for protein metabolism and is also an essential amino acid in birds' nutrition. Therefore, the birds have an absolute need for L-Arg and are highly dependent on its dietary supplementation (Miao et al, 2017). The Arg also serves as a substrate for the generation of creatine, polyamines, L-proline, various hormones, and nitric oxide (NO), which serves multiple roles within the immune system (Khajali and Wideman, 2010). It was reported that reduced oviduct weight and the number of small yellow follicles, also increased yolk color and immunoglobulin Y (IgY) content of the eggs by arginine supplementation of diet (17 mg L-Arg/kg diet) in compression to other treatment. In contrast, no effects were observed on laying performance, egg quality, and blood parameters (Yang et al., 2016). Liu et al (2014) have shown that an improvement in the innate and

acquired immune system of chickens fed with Argsupplemented diets and also challenged with Salmonella.

Several studies have shown that the positive impact of dietary arginine supplementation higher than the recommended levels on the performance of laying hens (NRC, 1994). Youssef et al. (2016) stated that Arg supplementation at a higher level than NRC could significantly increase egg production and egg mass during the whole experimental period. There was at least 6% of extra egg production over control. Fernandes et al. (2009) have suggested that dietary supplementation with Arg at higher levels than recommended for the starter phase may be necessary for improved muscle development in broilers. Also, Silva et al (2012), reported an increase in egg production and egg mass by L-Arg dietary supplementation. Dietary Arg requirements of broilers may not be adequate and enough to support

Please cite this article as Ahmad Azizi, Aliasghar Saki, Hossein Jahanian Najafabadi, Pouya Zamani & Zeinab Bardel. 2023. Digestible Arginine Requirements in Hy-Line W-36 Laying Hens: Effects on Performance, Egg Characteristics, and Plasma Parameters During 40 to 46 Weeks of Age. Poult. Sci. J. 11(1): 103-113.

maximal growth, immune function, avoid pulmonary hypertension at high altitudes, cold environmental conditions, disease challenges, and management conditions (Khajali and Wideman, 2010). Although that L-Arg can stimulate protein synthesis, this is demonstrated in some in vitro studies in chickens or pigs (Yuan et al., 2015). The L- Arg improved weight gain and feed intake, especially in growing chickens and pullets independent of genetic background (Lieboldt et al., 2015). Amino acids are major regulators of growth and protein metabolism and diets deficient in a specific limiting amino acid could lead to decreased protein accretion (Tesseraud et al., 1999). Therefore, this study managed to determine the requirement of Arg digestible in laving hens by regression equations and evaluate the effects of Arg on performance, blood parameters, and IGF-1 status.

Materials and Methods Birds and Housing

In this experiment, 150 laying hens of Hy-Line W-36 strains were allocated to 5 treatments, 5 replicates, and 6 hens in each. The experiment duration was carried out from 40 to 46 weeks of age.. The temperature was kept at around 18 to 22°C in the poultry house. The room humidity varied from 52 to 64%, and the lighting program was set to 16 hours of light and 8 hours of darkness during the experiment period.

Two weeks before the experiment (38 to 39 weeks), was as an adaptation period. The experimental diets (Table 1) were adjusted according to the nutritional by Hy-Line recommendation guided W-36 commercial layers (2015). Diets in mash form were offered ad libitum and birds had free access to water throughout the experiment. Experimental treatments were obtained by adding 0.81, 0.86, 0.91, 0.96, and of digestible Arg (98% of purity, Merck, 1.01% Germany). Dry matter, ash, crude protein, crude fiber, and crude fat content of raw materials in dietary treatments (corn, soybean meal, barley, and corn gluten) were measured according to the AOAC (2000). Feed intake was provided twice per day (morning and evening) the residual of replicate feed was recorded at the end of each week and replaced by a new diet. There was no mortality, therefore, weekly feed intake was recorded based on the day hen by the following formula (Harms and Racell, 2001):

Day hen = $7 \times$ number of alive hens

Means of feed intake for each hen per day = Weekly alive hens feed intake/day hens

Egg mass = egg weight \times production percentage

FCR = Feed intake/Egg mass

Also, the amino acid profile of the feed ingredients was determined by Near-Infrared Reflectance Spectroscopy (NIRS) and standardized ileum digestible amino acids estimated by coefficients of Degosa Evonik (Amino-Dat. 5).

Experimental diets

Table 1	. Basic	dietary cor	nponents along	with ch	emical ana	lysis	(as %)
---------	---------	-------------	----------------	---------	------------	-------	-------	---

Corn	46.56
Soybean meal	19.29
Corn gluten meal	4.60
Barley	13.00
Soybean oil	3.22
Dicalcium phosphate	2.34
Oyster shell Powder	10.00
Salt	0.40
Mineral supplement	0.25
Vitamin supplement	0.25
D, L-Methionine	0.09
Chemical analysis	
Metabolism energy (kcal/kg)	2822
Crude protein (%)	16.37
Calcium (%)	4.40
Available P (%)	0.55
Sodium (%)	0.18
Chlorine (%)	0.29
Lysine (%)	0.71
Methionine	0.36
Methionine + Cystine (%)	0.60
Threonine (%)	0.50
Tryptophan (%)	0.15
Arginine (%)	0.81
Valine (%)	0.69
Isoleucine (%)	0.59
Drovided nor by of the mineral supplements 70 mg of monganess (ovi	da) 60 mg of ging (gyidg) 60 mg of iron (gylfota) 8 mg of

Provided per kg of the mineral supplement: 70 mg of manganese (oxide), 60 mg of zinc (oxide), 60 mg of iron (sulfate), 8 mg of copper (sulfate), 1.1 mg of iodine (calcium iodate), 0.15 milligrams of cobalt and 0.25 milligrams of selenium. Each kilo of vitamin supplement: 10,000 units of vitamin A, 2500 international units of vitamin D₃, 20 international units of vitamin E, 3 milligrams of vitamin K₃, 2 mg thiamine, 5 mg of riboflavin, 12 mg of pantothenic acid, 40 mg niacin 5200 mg choline chloride, 5 mg Pyridoxine, 0.015 mg of cobalamin, 0.05 grams of biotin, 400 mg choline chloride, 0.75 mg of folic acid.

Eggs quality characteristics

To evaluate the egg characteristics, two eggs were selected randomly from each replicate on two final days of each week and transferred to the laboratory. Initially, the length and width of each egg were measured to calculate the shape index, i.e., (width/length) \times 100. To examine the characteristics of the eggshell, the shells were washed, then placed in an oven at 65°C for 24 hours, and weighed. The thickness of the shells was measured by a digital micrometer (INSIZE, Taiwan) with a precision of 0.001 mm (Harms and Russell, 2001), egg weight (EW), and albumen height (AH) were measured to calculate the Haugh Unit as the following formula. $HU = 100 \text{ Log} (AH + 7.75 - 1.7 \text{ EW}^{0.37}).$

The yolk color was recorded by the color spectrum of the Rosh unit. Further, characteristics such as yolk index (YI) and shell ratio (SR) were calculated from the raw data by the following formulas (Altan et al., 1998; Keener et al., 2006).

 $SR = shell weight/Egg weight \times 100$

 $YI = yolk height/yolk diameter \times 100$

Plasma Chemical Analysis

To evaluate the effects of amino acid levels of Arg on plasma biochemical parameters, at the end of the experiment (46 weeks), two hens from each replicate were randomly selected and blood samples were taken from their vein's wings. Heparin anticoagulant tubes were used to prevent coagulation of blood samples. Immediately after authorization, the plasma samples were centrifuged at $4472 \times g$ for 20 minutes and stored at -20°C until analysis. In this study, four parameters of plasma, including cholesterol, uric acid, globulin, and IGF-1, were recorded according to common colorimetric methods (Spectrophotometer).

Statistical analysis

This experiment was conducted in a completely randomized design (CRD) with 5 treatments, 5 replicates, and 6 hens in each. Production characteristics including egg weight, egg production, feed intake, egg mass, feed conversion ratio, body weight change and egg quality, and plasma parameters, containing cholesterol, uric acid. globulin, and IGF-1 concentrations were analyzed as repeated measures. Means of treatments with significant effects in the analysis of variance were compared based on Duncan at a significance level of 0.05. Orthogonal polynomial tests were also performed to evaluate the response curves. Digestible Arg requirements for the traits, with significant treatment effects and significant quadratic polynomial responses, were estimated by derivation of quadratic regression models. The data were analyzed by SAS (2008).

Results

Performance

The effect of experimental treatments on the performance of laying hens during the whole production period is presented in Table 2. No significant differences were found between treatments on feed intake, egg weight, and body weight change in the whole period (P > 0.05). In the analysis of feed intake, the linear, quadratic and cubic regressions were not significant (P > 0.05). There were no significant differences in body weight between different treatments in the whole period of production (P > 0.05). However, no response was shown in egg weight by dietary treatments. No significant differences were observed by analysis of the linear, quadratic and cubic regression during all weeks in egg weight (P > 0.05).

Table 2. Effect of dietary arginine levels on performance traits during 40 to 46 weeks of age

Dietary Arg level (%)	Feed intake (g/day)	Egg mass (g/day)	Egg production (%)	FCR ¹ (g/g)	Egg weight (g)	CBW ² (g/period)
0.81	93.90	47.37°	77.28°	1.941 ^a	61.15	0.15
0.86	93.09	54.34 ^a	89.18 ^a	1.712 ^b	60.96	0.16
0.91	95.27	50.60 ^b	82.72 ^b	1.863 ^a	61.08	0.13
0.96	94.66	50.06 ^b	81.77 ^b	1.880^{a}	61.15	0.14
1.01	94.99	50.58 ^b	82.65 ^b	1.864 ^a	61.10	0.15
SEM ³	1.512	0.452	0.81	0.028	0.608	0.040
<i>P</i> -value ⁴						
Treatment	0.845	< 0.0001	< 0.0001	0.0003	0.999	0.244
L. R	0.441	0.153	0.246	0.858	0.959	0.858
Q. R	0.938	< 0.0001	< 0.0001	0.018	0.919	0.018
C. R	0.674	< 0.0001	< 0.0001	0.0001	0.826	0.0001
Week	< 0.0001	< 0.0001	< 0.0001	< 0.0001	< 0.0001	< 0.0001
Treatment×Week	0.0022	< 0.0001	< 0.0001	0.0027	0.9088	0.0027

¹ FCR, Feed conversion ratio.

² CBW, Changes in body weight.

³ SEM, Standard error of means.

⁴ L. R, linear regression; Q. R, Quadratic regression; C. R, Cubic regression.

The highest and the lowest value of egg production and egg mass were found in 0.86% and 0.81% Arg treatments, respectively (P < 0.05). The use of 0.86% Arg significantly improved the feed conversion ratio. However, these differences were not considered due to the significant interaction of treatment \times week during the whole experimental period. Therefore, it should be statistical analysis separately in different weeks (Tables 3, 4, and 5). In

addition, significant quadratic and cubic regression equations were found in egg production (Table 6), egg mass, and feed conversion ratio (P < 0.05). The effect of treatment on egg production, egg mass, and feed conversion ratio were significant in the fourth, fifth, sixth, and seventh weeks. In the 4th week, egg production significantly decreased in a level of 0.81% Arg compared with other treatments.

Table 3. Effect	s of dietary	arginine level	s on egg producti	on (%) of	laying hens	in different w	veeks (40 to 46 weeks
-----------------	--------------	----------------	-------------------	-----------	-------------	----------------	-----------------------

Dietary Arg level (%)	Week 1	Week 2	Week 3	Week 4	Week 5	Week 6	Week 7
0.81	93.34	87.14	83.33	76.66 ^b	71.43°	64.28 ^c	64.76 ^c
0.86	91.91	89.52	82.85	86.66 ^a	90.95 ^a	90.47 ^a	91.90 ^a
0.91	91.91	87.62	84.76	85.71 ^a	80.95 ^b	74.76 ^b	73.33 ^b
0.96	86.19	90.47	82.85	84.76 ^a	78.09 ^b	75.23 ^b	74.76 ^b
1.01	90.00	91.43	83.33	84.28 ^a	80.47 ^b	74.28 ^b	74.76 ^b
SEM ¹	0.763	2.386	1.826	1.587	1.850	1.564	2.064
<i>P</i> -value ²							
Treatment	0.0781	0.6741	0.9440	0.0018	< 0.0001	< 0.0001	< 0.0001
L. R	0.0388	0.2215	0.9997	0.0152	0.3821	0.3476	0.6670
Q. R	0.4781	0.8333	0.7827	0.0021	0.0008	< 0.0001	< 0.0003
C. R	0.1621	0.7556	0.9992	0.0338	< 0.0001	< 0.0001	< 0.0001
a-c Different letters indicate	aignificant diff	anon and hater		Dum agm's tag	+(D < 0.05)		

Different letters indicate significant differences between groups in Duncan's test (P < 0.05).

¹ SEM, Standard error of means.

² L. R, linear regression; Q. R, Quadratic regression; C. R, Cubic regression.

Table 4.	Effects of di	ietary arginine l	evels on egg mass	(g/day) of lavir	ng hens in different	weeks (40 to 46 weeks)
14810 11	Billetto of a	ierai j'aiginnie i		(g, ddj) or idji		

Dietary Arg level (%)	Week 1	Week 2	Week 3	Week 4	Week 5	Week 6	Week 7
0.81	58.77	54.95	51.35	46.07 ^b	42.61 ^c	38.19 ^c	39.63°
0.86	58.40	55.84	50.26	52.41 ^a	54.66 ^a	53.03 ^a	55.78 ^a
0.91	58.50	54.42	52.31	51.96 ^a	48.69 ^b	44.27 ^b	44.10 ^b
0.96	54.64	56.90	50.85	50.87 ^a	47.06 ^b	44.91 ^b	45.14 ^b
1.01	57.09	57.80	50.68	51.05 ^a	48.54 ^b	43.95 ^b	44.89 ^b
SEM ¹	1.186	1.437	1.153	0.988	1.170	1.019	1.401
<i>P</i> -value ²							
Treatment	0.1211	0.4679	0.7615	0.0014	< 0.0001	< 0.0001	< 0.0001
L. R	0.0728	0.1527	0.8422	0.0138	0.2625	0.3042	0.9804
Q. R	0.7097	0.4757	0.7013	0.0022	0.0010	< 0.0001	< 0.0001
C. R	0.1353	0.8750	0.6162	0.0177	< 0.0001	< 0.0001	< 0.0001

^{a-c} Different letters indicate significant differences between groups in Duncan's test (P < 0.05).

¹ SEM, Standard error of means.

² L. R, linear regression; Q. R, Quadratic regression; C. R, Cubic regression.

Table 5.	Effects of	f dietary	arginine	levels o	n feed	conversion	ratio (g	g/g) o	f laying	hens in	different	weeks ((40 to	46
weeks)														

Dietary Arg level (%)	Week 1	Week 2	Week 3	Week 4	Week 5	Week 6	Week 7
0.81	1.700	1.815	1.965	2.011 ^a	1.955ª	2.123 ^a	2.011 ^a
0.86	1.651	1.722	1.852	1.790 ^b	1.733 ^b	1.688 ^c	1.590 ^b
0.91	1.703	1.833	1.824	1.864 ^{ab}	1.901 ^a	1.963 ^b	1.932 ^a
0.96	1.804	1.733	1.883	1.913 ^{ab}	1.950 ^a	1.970 ^b	1.946 ^a
1.01	1.721	1.705	1.924	1. 872 ^{ab}	1.892 ^a	1.971 ^b	1.934 ^a
SEM ¹	0.051	0.053	0.073	0.046	0.041	0.048	0.037
<i>P</i> -value ²							
Treatment	0.3120	0.3901	0.6814	0.0449	0.0076	< 0.0001	< 0.0001
L. R	0.2153	0.2295	0.9588	0.3473	0.5236	0.9131	0.0990
Q. R	0.9141	0.6912	0.2037	0.0632	0.2170	0.0029	0.0023
C. R	0.0963	0.4622	0.4990	0.0178	< 0.0011	< 0.0001	< 0.0001

^{a-c} Different letters indicate significant differences between groups in Duncan's test (P < 0.05).

¹ SEM, Standard error of means.

² L. R, linear regression; Q. R, Quadratic regression; C. R, Cubic regression.

In 5, 6, and 7 weeks, egg production significantly decreased with 0.81% Arg, but reverse trend was noted in 0.86% Arg treatment. The egg mass significantly decreased with the level of 0.81% Arg in week 4, but there were no significant differences between the other treatments. In 5, 6, and 7 weeks, egg mass significantly decreased with the level of

0.81% Arg and increased significantly with 0.86% Arg. In the 4th and 6th weeks, the feed conversion ratio significantly increased by 0.81 and decreased by 0.86% Arg. In 5 and 7 weeks, FCR decreased significantly with a level of 0.86% Arg, but no significant differences were shown by other treatments.

	Table 6.	Effect of	dietary	arginine	levels on	the egg	quality	of laying	hens	(from 4	40 to 4	6 weeks	of age	ļ
--	----------	-----------	---------	----------	-----------	---------	---------	-----------	------	---------	---------	---------	--------	---

Dietary Arg lavel (%)	Shape index	Yolk index	Yolk color	Haugh unit	Shell thickness	Shell ratio
Dietal y Aig level (70)	(%)	(%)	(Roche)		(mm)	(%)
0.81	75.67	44.80	6.82	83.83	0.470	8.84
0.86	75.28	45.59	6.82	82.89	0.481	8.73
0.91	75.08	45.37	7.10	83.64	0.502	9.05
0.96	75.94	45.47	6.75	84.03	0.473	9.00
1.01	73.91	45.89	6.78	83.57	0.484	9.33
SEM ²	0.883	0.660	0.125	0.689	0.007	0.177
<i>P</i> -value ²						
Treatment	0.5481	0.8268	0.3155	0.8136	0.1754	0.2079
L. R	0.3178	0.3317	0.6627	0.7873	0.9563	0.0400
Q. R	0.5057	0.8701	0.2332	0.8258	0.1162	0.4541
C. R	0.2839	0.5315	0.8029	0.2567	0.3306	0.9440
Week	0.6030	< 0.0001	< 0.0001	< 0.0001	< 0.0001	0.0281
Treatment×Week	0.3762	0.2862	< 0.0001	0.1708	0.0036	0.0024

^{a-c} Different letters indicate significant differences between groups in Duncan's test (P < 0.05).

¹ SEM, Standard error of means.

² L. R, linear regression; Q. R, Quadratic regression; C. R, Cubic regression.

Egg characteristics

The high level of dietary Arg (% 0.96) leads to an increased yolk color (Table 7). The analysis of linear, quadratic, and cubic regression relationships for all egg characteristics was not significant in the experimental whole period (P > 0.05).

Blood parameters

The effect of treatments on, globulin, IGF-1, uric acid,

and cholesterol of birds are represented in Table 8. No significant effect of Arg treatments was found on plasma globulin, uric acid, cholesterol, and IGF-1 by linear, quadratic, and cubic regression. In addition, a significant increase was found in plasma IGF1 by 0.91% Arg in control significant decrease was shown by 0.81 Arg in this respect, in comparison to other treatments (P < 0.05).



Figure 1. Dietary requirement of arginine (%) to maximize egg production.

Table 7. Effec	ots of dietar	y arginine l	evels on Yo	lk color, Shel	l thickness, a	and Shell rat	tio in differe	nt weeks (40 t	to 46 weeks).			
	Yolk	color (Roch	le)			Shell thick	cness (mm)			Shell ra	tio (%)	
Dietary Arg level (%)	Week40	Week42	Week44	Week46	Week40	Week42	Week44	Week46	Week40	Week42	Week44	Week46
0.81	6.30	6.70^{bc}	7.50	$6.80^{\rm abc}$	0.480	0.477^{a}	0.50^{0ab}	0.463^{b}	8.58	8.97 ^a	8.95	8.87^{ab}
0.86	6.00	7.20^{ab}	7.00	7.10^{ab}	0.491	0.496^{a}	0.541 ^a	0.542^{a}	9.03	8.25 ^b	9.23	8.42 ^b
0.91	6.10	7.60^{ab}	7.20	7.50^{a}	0.495	0.497^{a}	0.532^{a}	0.534^{a}	8.87	9.12 ^a	8.92	9.29^{ab}
0.96	5.70	7.70^{a}	7.10	6.50^{bc}	0.488	0.465^{b}	0.504^{ab}	0.501^{ab}	9.02	8.84^{ab}	8.24	9.88^{a}
1.01	5.60	5.60°	7.50	6.20°	0.519	0.481^{a}	0.483^{b}	0.484^{b}	8.78	9.31 ^a	9.14	10.07^{a}
SEM^{1}	0.125	0.154	0.223	0.250	0.009	0.012	0.015	0.015	0.162	0.200	0.320	0.403
P-value ²												
Treatment	0.2121	0.0003	0.4020	0.0141	0.1080	0.0329	0.0472	0.0472	0.2870	0.0151	0.2487	0.0476
L. R	0.0197	0.0001>	0.8890	0.0346	0.1517	0.2436	0.0570	0.9634	0.4506	0.0604	0.5652	0.0065
Q. R	0.9010	0.0723	0.0881	0.0119	0.1970	0.2722	0.0260	0.0203	0.0892	0.1145	0.4771	0.5097
C. R	0.8830	0.8403	0.7802	0.4585	0.0463	0.1039	0.2314	0.5259	0.6764	0.1981	0.0442	0.1959
^{a-c} Different le	tters indicat	e significan	nt difference	s between gro	oups in Dunc	an's test (P	< 0.05).					
¹ SEM, Standa	urd error of 1	means.										
2 I D linearr		D O O	notio regrege	U D U	in ramaration	2						

L. R, linear regression; Q. R, Quadratic regression; C. R, Cubic regression.

108

Dietary Arg level (%)	IGF-1	Globulin	Uric acid	Cholesterol
	1 (µg/dL)	(ng/ mL)	(mg/dL)	(mg/dL)
0.81	30.75 ^c	2.34	8.05	100.70
0.86	36.86 ^{bc}	2.22	5.02	135.70
0.91	55.18 ^a	2.36	5.00	131.00
0.96	38.85 ^{bc}	3.02	4.13	175.60
1.01	51.08 ^{ab}	2.34	4.45	123.30
SEM ²	5.416	0.348	0.949	26.062
<i>P</i> -value ³				
Treatment	0.0322	0.5125	0.0576	0.3880
L. R	0.0173	0.4757	0.0139	0.3141
Q. R	0.4275	0.6499	0.1073	0.2135
C. R	0.4550	0.1615	0.5512	0.4956

Table 8. Effect of arginine amino acid level on some plasma parameters

^{a-c} Different letters indicate significant differences between groups in Duncan's test (P < 0.05).

¹ IGF-1, Insulin-like growth factor.

² SEM, Standard error of means.

³ L. R, linear regression; Q. R, Quadratic regression; C. R, Cubic regression.

Table 9. The quadratic regression equation and level of required arginine

Traits	Quadratic regression equation	Estimated Arginine Requirement (dig %) ¹
Egg Production	Y= -312.10 + 866.28X - 472.32X2	0.917
Egg mass	Y= -182.23 + 510.49X - 278.15X2	0.917
FCR ²	Y= 8.33 - 14.35X + 7.90X2	0.908
IGF-1 ³	Y=75.98 + 33.073X - 14.04X2	0.970

¹Requirements base Digestible

2 Feed conversion ratio.

³ Insulin-like growth factor 1

Digestible Arg requirement

Estimation of digestible Arg requirement was carried out by using the quadratic regression method. The equivalence of the quadratic regression and the required levels of Arg are shown in Table 9. The estimated levels of Arg requirements for egg production, egg mass, feed conversion ratio and IGF-1 were 0.917, 0.917, 0.908, and 0.970%, respectively (Figures 1, 2, 3, and 4 on the curves, respectively).



Figure 2. Dietary requirement of arginine (%) to maximize egg mass.



Figure 3. Dietary requirement of arginine (%) to minimize feed conversion ratio.



Figure 4. Dietary requirement of arginine (%) to maximize blood plasma IGF-1 concentration.

Discussion

Poultry performance is affected by the essential amino acids. However, high levels of any of the amino acids may have a negative effect on performance and the antagonism between amino acids such as Arg and lysine could be observed (D'mello, 2003). Analysis of data on feed conversion ratio did not show any significant differences between experimental treatments during the whole experimental period. In contrast in the final weeks, significantly different levels of Arg influenced the feed conversion ratio, and these results were confirmed by Lieboldt *et al.*, (2016) and Jahanian and Khalifeh-Gholi, (2018) who have shown negative

effects of dietary Arg reduction on feed conversion ratio.

Analysis of data on the percentage of production in the whole experimental period did not show any differences among treatments. But there were significant differences in the increase of production in the fourth, fifth, sixth, and seventh weeks by % 0.86 of Arg. Increasing production in some research may be due to the improvement of the amino acid balance in ration and Arg intake of chickens for maximum production.

On the other hand, an increase in the level of Arg may increase the secretion of the LH hormone, followed by an increase in egg production (Basiouni, 2009). In contrast higher levels of Arg resulted in a reduction in the percentage of production. Increased egg production by increasing Arg levels could be obtained but a higher Arg level more than the requirement status could lead to decrease egg production this may be related to Arg and lysine antagonism. Also, the Arg amino acid of the Nmolecule is highly integrated with the structure. hence the poultry require a lot of energy for the metabolism process and the removal of excess Arg. Further increases in the level of Arg did not increase the amount of egg production and were consistent with the results of Najib and Basiouni (2004) and Basiouni et al., (2006). It has been reported that excess Arg in the diet increases uric acid excretion. On the other hand, a molecule of glycine is excreted with the removal of any uric acid molecule. The glycine amino acid is synthesized in the poultry body to a limited extent, and when poultry encounters an excess of uric acid, it does not meet the needs of tissues and organs (Corzo et al., 2004). Following this, may decrease the percentage of egg production.

Analysis of data on average egg weight in different weeks and the whole experiment period did not show any significant differences among experimental treatments. The results of the mean egg weight in response to the Arg supplement in this study were compared with previous studies by Carvalho et al. (2012), Yuan et al. (2015), and Youssef et al. (2016). In an analysis of data on the average egg mass over the whole trial period, there were no significant differences between experimental treatments. According to previous research results, higher levels of Arg reduce the average egg mass. Achieving these results was not unexpected. Because the egg mass depends on the percentage of production and the weight of the egg production. An analysis of feed intake data did not show any significant differences among experimental treatments during the whole period of study. The results of this study on feed intake are consistent with the results by Rubin et al. (2007), Carvalho et al. (2012), and Youssef et al. (2016).

There is limited literature considering the effects of Arg acid on egg quality. Analysis of data on quality traits did not show a significant difference in the characteristics of shape index, yolk index, yolk color, eggshell, Haugh unit, and shell weight. According to Carvalho *et al.* (2012) and Yang *et al.* (2016) results, increasing the level of Arg in laying hens did not have any significant effect on the egg quality

However, Carvalho *et al*, (2015) have reported that increasing levels of digestible lysine and Arg lead to reduced eggshell quality and albumen solids, respectively. Yang *et al*. (2016) also indicated that increased levels of dietary Arg increased the yolk color. These results are consistent with the results of the present study. Yuan *et al.* (2015) observed no significant differences in egg characteristics by increasing the Arg level in the diet, but an exception was shown by significantly improved Haugh unit. De Lima and da Silva (2007), Silva *et al.* (2012), and Youssef *et al.* (2016) no significant differences were recognized in the internal egg quality characteristics by increasing Arg levels. Results of the present study have shown that increasing the levels of Arg leads to an increase in the shell thickness of the eggs, by improving mineral deposition in the bone and influencing the metabolism of the minerals Carlisle

(1986); Visser and Hoekman (1994). In the present study, no significant differences were indicated in plasma cholesterol by increasing dietary Arg levels in laying hens, However, the study of Emadi et al. (2010) and Fouad et al. (2013) were inconsistent with our results. They have stated that dietary Arg leads to decrease plasma triglyceride and total cholesterol concentrations in broilers fed Argsupplemented diets. Fouad et al. (2013) have shown that the Arg mechanism effect on plasma cholesterol is still not well known and requires further studies. There are very few studies that have considered the effects of Arg on IGF-1 plasma levels. Arg is considered a secretagogue, increasing the level of IGF-A in the blood (Silva et al., 2012). In this experiment, analyzing data on plasma IGF-1 levels showed that increasing the level of Arg in the diet increases the level of this parameter in the blood plasma. These results agree with the results of the study by Yu et al., (2018). They have indicated that increased levels of Arg in the plasma increase the secretion of the hormone IGF-1 and increase its level in the blood plasma. It has been reported that increasing the amount of Arg in the feed increases the synthesis of ornithine. Turnitin increases the secretion of the growth hormone by stimulating the pituitary gland (Isidori et al. 1981; Sugino et al. 2008). Analysis of data on plasma globulin levels has shown that increased levels of dietary Arg had no significant effect on plasma globulin levels.

The dietary requirement of L-Arg, noted by the NRC is not sufficient to maximize the performance of white and brown-egg-laying strains (Yuan *et al.*, 2015). Schutte *et al.*, (1994) have stated that the estimation of nutrient requirements may depend on the biological response and quantity of the requirement. In this study, the quadratic equations showed that digestible Arg requirement for maximum egg mass, production percentage, and insulin-like level of hormone in blood plasma, was 0.917, 0.917, and 0.970% respectively, and the best level for the feed conversion ratio was estimated at 0.908 %. Yuan *et al.*, (2015) have shown that levels higher than the recommended of L-Arg increased the liver protein synthesis rate and protein gain rate of laying hens.

Conclusion

The present study has found that the content of digestible Arg in the diet may be helpful to improve the secretion of IGF-1, feed conversion ratio, egg production, egg mass, and yolk color of laying hens from 40 to 46 weeks of age. The quadratic equations analysis showed that the optimum levels of digestible Arg for improvement of egg mass, egg production

References

- Altan O, Oguz I & Akbas Y. 1998. Effects of selection for high body weight and age of hen on egg characteristics in Japanese quail (*cotumix cotumix japonica*). Journal of Veterinary and Animal Science, 22: 464-473.
- AOAC. 2000. Official Methods of Analysis, 17th ed. AOAC International, Gaithersburg, MD.
- Basiouni G, Najib H, Zaki MM & Al-Ankari AS. 2006. Influence of extra supplementation with arginine and lysine on overall performance, ovarian activities and humoral immune response in local Saudi hens. International Journal of Poultry Science, 5(5): 441-448. DOI: 10.3923/ijps.2006.441.448
- Basiouni GF. 2009. The effect of feeding an extra amount of arginine to local Saudi hens on luteinizing hormone secretion. Journal of Biological Sciences, 9(6): 617-620. DOI: 10.3923/jbs.2009.617.620
- Carlisle EM .1986. Silicon as an essential trace element in animal nutrition, in: Evered, D. and O'Connor, M. (Eds) Silicon Biochemistry. Ciba Foundation Symposium, 121: 123-139.
- Carvalho FB, Stringhini JH, Matos MS, Café MB, Leandro NSM, Gomes NA, Santana ES & Jardim Filho RDM. 2015. Egg quality of hens fed different digestible lysine and arginine levels. Brazilian Journal of Poultry Science, 17(1): 63-68. DOI: 10.1590/1516-635x170163-68
- Carvalho FB, Stringhini JH, Matos MS, Jardim F, Cafe MB, Leandro NM & Andrade MA. 2012. Performance and nitrogen balance of laying hens fed increasing levels of digestible lysine and arginine. Revista Brasileira de Zootecnia, 41(10): 2183-2188. DOI: 10.1590/S1516-35982012001000007
- Corzo A, Kidd MT, Burnham DJ & Kerr BJ. 2004. Dietary glycine needs of broiler chicks. Poultry Science, 83(8): 1382-1384. DOI: 10.1093/ps/83.8.1382
- De Lima MR & da Silva JHV. 2007. Efeito da relação lisina: arginina digestível sobre o desempenho de poedeiras comercias no período de postura. Acta Veterinária Brasílica, 1(4): 118-124. DOI: 10.21708/avb.2007.1.4.519
- D'mello JPF. 2003. Amino acids as multifunctional molecules, in: D'mello, J.P.F. Jr. (Ed.). Amino

percentage, plasma IGF-1, and feed conversion ratio, were 0.917, 0.917, 0.970, and 0.908 % respectively.

Acknowledgments

The authors thank Department of Animal Science at Bu-Ali Sina University for excellent scientific collaboration and financial support.

Acids in Animal Nutrition. Pages, 87-101 (CABI Publishing).

- Emadi M, Kaveh K, Bejo MH, Ideris A, Jahanshiri F, Ivan M & Alimon RA. 2010. Growth performance and blood parameters as influenced by different levels of dietary arginine in broiler chickens. Journal of Animal and Veterinary Advances, 9(1): 70-74. DOI: 10.3923/javaa.2010.70.74
- Fernandes JIM, Murakami AE, Martins EN, Sakamoto MI & Garcia ERM. 2009. Effect of arginine on the development of the pectoralis muscle and the diameter and the protein: deoxyribonucleic acid rate of its skeletal myofibers in broilers. Poultry Science, 88(7): 1399-1406. DOI: 10.3382/ps.2008-00214
- Fouad AM, El-Senousey HK, Yang XJ & Yao JH. 2013. Dietary L-arginine supplementation reduces abdominal fat content by modulating lipid metabolism in broiler chickens. Animal: An International Journal of Animal Bioscience, 7(8): 1239-1245. DOI: 10.1017/S1751731113000347
- Harms RH & Russell GB. 2001. Evaluation of valine requirement of the commercial layer using a cornsoybean meal basal diet. Poultry Science, 80(2): 215-218. DOI: 10.1093/ps/80.2.215
- Isidori A, Lo Monaco A & Cappa M. 1981. A study of growth hormone release in man after oral administration of amino acids. Current Medical Research and Opinion, 7(7): 475-481. DOI: 10.1185/03007998109114287
- Jahanian R & Khalifeh-Gholi M. 2018. Marginal deficiencies of dietary arginine and methionine could suppress growth performance and immunological responses in broiler chickens. Journal of Animal Physiology and Animal Nutrition, 102(1): e11-e20. DOI: 10.1111/jpn.12695
- Keener KM, McAvoy KC, Foegeding JB, Curtis PA, Anderson KE & Osborne JA. 2006. Effect of testing temperature on internal egg quality measurements. Poultry Science, 85: 550-555. DOI: 10.1093/ps/85.3.550
- Khajali F & Wideman RF. 2010. Dietary arginine: metabolic, environmental, immunological and physiological interrelationships. World's Poultry Science Journal, 66(4): 751-766. DOI: 10.1017/S0043933910000711
- Lieboldt MA, Frahm J, Halle I, Görs S, Schrader L,

Weigend S, Preisinger R, Metges CC, Breves G, & Dänicke S. 2016. Metabolic and clinical response to Escherichia coli lipopolysaccharide in layer pullets of different genetic backgrounds supplied with graded dietary L-arginine. Poultry Science, 95(3): 595-611. DOI: 10.3382/ps/pev359

- Lieboldt MA, Halle I, Frahm J, Schrader L, Weigend S, Preisinger R & Dänicke S. 2015. Effects of long-term graded L-arginine supply on growth development, egg laying and egg quality in four genetically diverse purebred layer lines. The Journal of Poultry Science, 53(1): 8-21. DOI: 10.2141/jpsa.0150067
- Liu X, Byrd JA, Farnell M & Ruiz-Feria CA. 2014. Arginine and vitamin E improve the immune response after a Salmonella challenge in broiler chicks. Poultry Science, 93(4): 882-890. DOI: 10.3382/ps.2013-03723
- Miao LP, Yuan C, Dong XY, Zhang XY, Zhou MY & Zou XT. 2017. Effects of dietary L-arginine levels on small intestine protein turnover and the expression of genes related to protein synthesis and proteolysis of layers. Poultry Science, 96(6): 1800-1808. DOI: 10.3382/ps/pew471
- Najib H & Basiouni GD. 2004. Determination of the nutritional requirements of the Local Saudi chickens. 1. Effect of Arginine inclusion, in excess of the leghorn requirement, on performance of the Local Saudi chickens. Scientific Journal of King Faisal University Basic and Applied Sciences, 5: 121-144.
- NRC (National Research Council). 1994. Nutrient Requirements of Poultry. 9th Rev. Ed. National Academy Press. Washington, DC. 176 Pages.
- Rubin LL, Canal CW, Ribeiro AL, Kessler A, Silva I, Trevizan L, Viola T, Raber M, Gonçalves TA & Krás R. 2007. Effects of methionine and arginine dietary levels on the immunity of broiler chickens submitted to immunological stimuli. Brazilian Journal of Poultry Science, 9(4): 241-247. DOI: 10.1590/S1516-635X2007000400006
- SAS (Statistical Analysis System). 2008. SAS/STAT® 9.2. User's Guide. SAS Institute Inc. Cary, North Carolina.
- Schutte JB, De Jong J & Bertram HL. 1994. Requirement of the laying hen for sulfur amino acids. Poultry Science, 73(2): 274-280. DOI: 10.3382/ps.0730274

- Silva LMGS, Murakami AE, Fernandes JIM, Dalla Rosa D & Urgnani JF. 2012. Effects of dietary arginine supplementation on broiler breeder egg production and hatchability. Brazilian Journal of Poultry Science, 14(4): 267-273. DOI: 10.1590/S1516-635X2012000400006
- Sugino T, Shirai T, Kajimoto Y & Kajimoto O. 2008. L-ornithine supplementation attenuates physical fatigue in healthy volunteers by modulating lipid and amino acid metabolism. Nutrition Research, 28(11): 738-743. DOI: 10.1016/j.nutres.2008.08.008
- Tesseraud S, Le Bihan-Duval E, Peresson R, Michel J & Chagneau AM. 1999. Response of chick lines selected on carcass quality to dietary lysine supply: live performance and muscle development. Poultry Science, 78(1): 80-84. DOI: 10.1093/ps/78.1.80
- Visser JJ & Hoekman K. 1994. Arginine supplementation in the prevention and treatment of osteoporosis. Medical Hypotheses, 43(5): 339-342. DOI: 10.1016/0306-9877(94)90113-9
- Yang H, Ju X, Wang Z, Yang Z, Lu J & Wang W. 2016. Effects of arginine supplementation on organ development, egg quality, serum biochemical parameters, and immune status of laying hens. Brazilian Journal of Poultry Science, 18(1): 181-186. DOI: 10.1590/1516-635x1801181-186
- Youssef SF, Badawy MI & Abd El-Halim HAH. 2016. Effect of l-arginine supplementation on productive, reproductive performance, immune response and gene expression in two local chicken strains: 2-responses of offspring. Egyptian Poultry Science Journal, 36: 825 -839.
- Yu J, Yang H, Wang Z, Dai H, Xu L & Ling C. 2018. Effects of arginine on the growth performance, hormones, digestive organ development and intestinal morphology in the early growth stage of layer chickens. Italian Journal of Animal Science, 17(4): 1077-1082. DOI: 10.1080/1828051X.2018.1434692
- Yuan C, Li JM, Ding Y, He Q, Yan HX, Lu JJ & Zou XT. 2015. Estimation of L-arginine requirement for Xinyang Black laying hens from 33 to 45 weeks of age. Journal of Applied Poultry Research, 24(4): 463-469. DOI: 10.3382/japr/pfv049